Using Available Information in the Assessment of Diagnostic Protocols

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**Abstract**

A new binary screening or diagnostic test may be combined sequentially with an existing test to either improve the overall diagnostic performance or to reduce the overall cost while maintaining performance. If we assume that the tests produce binary results there are two protocol choices. A believe the positive protocol produces a positive result for all combinations of the existing and new test results except when both tests are negative while a believe the negative protocol produces a positive result only when both tests are positive. In practice the tests may be applied in sequence so we refer to an add-on plan as one in which the existing test is used first with the new test applied to only a subset of subjects depending on the protocol in use. In a triage plan, the new test is used on all subjects and the results determine who receives the existing test. Therefore, with two binary tests, a total of four sequential diagnostic protocols are possible.

Sequential protocols are used in a wide variety of settings including computed tomography as an add-on test to pelvic ultra sonography in a believe the positive protocol for the diagnosis of suspected appendicitis in children in Peña et al. [1], and high-risk human papillama virus testing as a triage test to cytological screening in a believe the negative protocol for precancers cervical lesions in Kotaniemi-Talonen et al. [2]. Although we refer to existing and new tests either could itself be a protocol combining multiple tests as long as the final result is dichotomous. For example, Gyselaers et al. [3] designed a screening protocol for trisomy 21 detection in the first trimester using a combination of age and serum test results as a triage to identify intermediate risk cases for advanced ultrasound scanning.

Our purpose is to demonstrate the value, expressed in terms of improved precision of important estimators, of using available information on the performance of the existing test, in combination with data from an assessment study of the existing and new tests to assess the properties of a combined protocol and compare the protocol to the existing test.

The available information or baseline data takes the form of a previous case-control study used to estimate the sensitivity and specificity of the existing test [4,5]. Initially, we confine ourselves to the setting where the assessment study is also a case-control study using a paired design (i.e. both tests are applied to all subjects). We assume that verification via a gold standard is available for both the assessment study and the baseline data. Later we discuss the use of cohort studies, where the numbers of diseased and non-diseased subjects are not fixed, and non-paired sequential assessment studies, where the new test is given to only a subset of subjects.

We make a key assumption that the diagnostic properties (sensitivity and specificity) of the existing test are the same in the baseline and assessment studies. Practically, this means selecting a source of baseline data with a population and test implementation as similar as possible to the assessment study. Depending on the disease under study, important population factors might include age- and sex-distributions and disease strain or severity in the cases. We derive a formal test of this hypothesis. If we reject the hypothesis then it is likely not appropriate to incorporate the baseline data into the analysis and either a new source of baseline data should be found or a standard analysis of the assessment study should be conducted. However, if we fail to reject the hypothesis, we will proceed with incorporating the available baseline data into the analysis of the combined protocol.

To assess the properties of the new protocol, we use maximum likelihood estimation for the sensitivity and specificity and asymptotic properties of the likelihood to find approximate standard errors. The comparison of the new protocol versus the existing test is made via likelihood ratios. These capture the trade-off between sensitivity and specificity of the new protocol versus the existing test [6-8]. Informally,
the positive (negative) likelihood ratio represents how many times more often a subject with the disease is expected to have a positive (negative) test result than one without the disease. The comparison between likelihood ratios is equivalent to comparing the predictive values of the two tests, when applied to the same population of subjects.

Likelihood ratios have previously been used by Nefuentes and del Castillo [9] to compare and test for differences between two binary tests and by Macaskill et al. [7] to compare a combined test with one of its components. Muwonge et al. [10] considered the sensitivities, specificities, and likelihood ratios for combinations of screening tests for cervical cancer. Bossuyt et al. [11] discussed the various ways of combining a new with an existing diagnostic test and Hayen et al. [8] discussed estimation of the likelihood ratios for a full paired design. Ahmed et al. [12] and Ahmed et al. [13] considered three logic rules for combining two tests based on continuous markers and compared them in terms of the false positive rate, the maximum receiver operating characteristic curve, and overall cost. While sequential tests are relatively common in medicine, we are not aware of previous work on incorporating baseline data. The use of baseline data is more common in industrial applications [14,15].

The remainder of the article is organized as follows. In the Methods section we define notation and formulate the sensitivity and specificity of the combined protocols and the likelihood ratios used to compare the believe the positive or believe the negative protocol versus the existing test. We also derive the relevant asymptotic variances and discuss estimation. In the Application section we apply the methods to an example of the use of mammography and sonography/ultrasound for breast cancer detection in symptomatic women. Next we conduct a factorial experiment to study the gains in precision across a wide range of sensitivities, specificities and study sizes. Finally, we conclude with a discussion.

Methods

The results of a paired case-control assessment study of two binary diagnostic tests with verification via a gold standard can be summarized in two 2×2 tables, one for diseased subjects and one for non-diseased subjects. Let E and N be binary random variables representing the result (1 for a positive result, 0 for a negative result) of the existing and new tests, respectively. Let D be a binary random variable representing the disease status (1 for the diseased, 0 for the non-diseased). We denote the cell frequencies for the assessment study by e, n, d, with e, n, d = 0, 1. Let n E be the total number of diseased subjects in the assessment study (n E = n E 0 + n E 1 + n N 0 + n N 1), n E 0 be the total non-diseased subjects, and n = n E + n N be the total sample size.

We define $\pi^E = Pr(E = e | D = 0)$ to be the joint probability of the results of the existing and new tests conditional on disease status. Using the convention of a + indicating marginalization or summation over that subscript, the sensitivity and specificity of the existing test from the paired assessment study are given by:

$$sens = Pr(E = 1 | D = 1) = \pi^E = 1 - \pi^N$$

$$spec = Pr(E = 0 | D = 0) = 1 - \pi^N$$

Additionally, we assume we have available baseline information about the performance of the existing test. We denote these data by $m^E$, e, d, with e, d = 0, 1. The total number of diseased subjects in the baseline is denoted by m, the number of non-diseased subjects by m N, and the total sample size for the baseline study by m = m + m N. An example of data of this type is given in Table 1 which will be discussed in detail in the.

| Table 1: Results of mammogram (Mx) and ultrasound (US) screening from the parallel assessment study of Houssami et al. [16] and mammogram baseline data from Kavanagh et al. [17]. |

<table>
<thead>
<tr>
<th>Combination</th>
<th>Breast cancer cases</th>
<th>Non breast cancer controls</th>
<th>Baseline data</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mx</td>
<td>148</td>
<td>34</td>
<td>182</td>
</tr>
<tr>
<td>Mx</td>
<td>8</td>
<td>21</td>
<td>29</td>
</tr>
<tr>
<td>Mx</td>
<td>98</td>
<td>1062</td>
<td></td>
</tr>
<tr>
<td>Mx</td>
<td>40</td>
<td>9208</td>
<td></td>
</tr>
<tr>
<td>Total</td>
<td>196</td>
<td>44</td>
<td>240</td>
</tr>
<tr>
<td>Total</td>
<td>138</td>
<td>10270</td>
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off between specificity and sensitivity does not necessarily translate into a trade-off between the two predictive values. It is possible for the believe the positive protocol to have higher positive and negative predictive values than the existing test. However, it is well known that the predictive values of a test depend on the accuracy of the test and the disease prevalence [7]. Therefore, a direct comparison between the predictive values of two tests is not recommended, as the results of the comparison cannot be extended beyond the population on which the assessment study was conducted.

Macaskill et al. [7] and Hayen et al. [8] propose comparing the new protocol with the existing test by looking at the logarithm of the ratio of their corresponding positive and negative likelihood ratios. The likelihood ratios of the two tests can be expressed in terms of the conditional probabilities defined in Equation (3) as follows

\[
LR^+_E = \frac{sens}{1 - spec} = \frac{\pi^+_1}{\pi^-_1}; \quad LR^-_E = \frac{1 - sens}{spec} = \frac{1 - \pi^+_1}{1 - \pi^-_1}
\]

\[
LR^+_UN = \frac{\pi^+_1 + \pi^+_2}{\pi^+_1 + \pi^-_2}; \quad LR^-_UN = \frac{\pi^-_1 + \pi^-_2}{\pi^-_1 + \pi^+_2}
\]

Marshall [6] shows that comparing the predictive values of two tests is equivalent to comparing the likelihood ratios of these tests, for fixed prevalence. That is, for fixed prevalence,

\[
PV^+_UN \geq PV^+_E \quad \text{if and only if} \quad LR^+_UN \geq LR^+_E
\]

and

\[
PV^-_UN \geq PV^-_E \quad \text{if and only if} \quad LR^-_UN \leq LR^-_E
\]

However, since the likelihood ratios do not depend on the disease prevalence we can compare the estimates of the likelihood ratios from the assessment study directly. This comparison translates into a comparison between the predictive values of the tests when applied to the same population.

The expressions for the logarithm of the ratios of the positive and negative likelihood ratios for the believe the positive protocol, denoted here \(\phi_E^{+\text{UN}}\) and \(\phi_E^{-\text{UN}}\), respectively, in terms of the conditional probabilities, are as follows:

\[
\phi_E^{+\text{UN}} = \ln \left( \frac{LR^+_UN}{LR^+_E} \right) = \ln[(\pi^+_1 + \pi^+_2)/(\pi^+_1 + \pi^-_2)] - \ln[(\pi^+_1 + \pi^+_2)/(\pi^+_1 + \pi^-_2)]
\]

(6)

\[
\phi_E^{-\text{UN}} = \ln \left( \frac{LR^-_UN}{LR^-_E} \right) = \ln[(1 - \pi^+_1 + \pi^-_2)/(1 - \pi^+_1 + \pi^-_2)] - \ln[(1 - \pi^+_1 + \pi^-_2)/(1 - \pi^+_1 + \pi^-_2)]
\]

(7)

If the believe the positive protocol is better than the existing test in terms of both the positive and negative predictive values then \(\phi_E^{+\text{UN}} > 0\) and \(\phi_E^{-\text{UN}} < 0\). However, it might be that the new protocol is better than the existing one in terms of the positive predictive value, but worse in terms of the negative predictive value, or the other way around.

**Analysis for the believe the positive protocol**

In this section, our goal is to quantify the gain in precision for the estimators of sensitivity and specificity of the believe the positive protocol, and of the log-ratios for both likelihood ratios, i.e., \(\phi_E^{+\text{UN}}\) and \(\phi_E^{-\text{UN}}\) that comes from incorporating the available baseline information into the estimation procedure.

Assuming independence between the baseline and assessment data, we can write the likelihood function as \(L \propto L_B \times L_E\), where \(L_B\) is the baseline likelihood

\[
L_B = (\pi^+_1)^{m_1} (1 - \pi^+_1)^{n_1-m_1} \times (\pi^-_1)^{m_2} (1 - \pi^-_1)^{n_2-m_2}
\]

and \(L_E\) is the assessment study likelihood

\[
L_E = (\pi^+_2)^{m_1} (1 - \pi^+_2)^{n_1-m_1} \times (\pi^-_2)^{m_2} (1 - \pi^-_2)^{n_2-m_2}
\]

(8)

Note that here we assume that the sensitivity (\(\pi^+_1\)) and specificity (\(1-\pi^-_1\)) of the existing test are the same in the baseline and assessment studies. This assumption should be tested as outlined in Appendix A of the Supplementary Materials. Using the combined likelihood function, the maximum likelihood estimates (MLEs) of the four conditional probabilities are

\[
\hat{\pi}^+_1 = \frac{m_1 + n_0}{m_1 + n_1}; \quad \hat{\pi}^-_1 = \frac{n_1 - m_0}{m_1 + n_1}, \quad \text{for} \quad D = 0,1.
\]

(9)

The MLEs of the sensitivity and specificity of the believe the positive protocol, and of the log-ratios of the likelihood ratios are obtained by substituting the MLEs given in Equation (10) into Equations (4-7).

The asymptotic variances of the estimators of \(s_{\text{UN}}\), \(s_{\text{EU}}\), \(\phi_E^{+\text{UN}}\), and \(\phi_E^{-\text{UN}}\) are found using the delta method (see the Appendix B of the Supplementary Materials for complete derivations). Given the numbers of diseased and non-diseased subjects in the assessment study and the baseline data, the asymptotic variances for the estimators of \(s_{\text{UN}}\) and \(s_{\text{EU}}\) are

\[
\text{var}[\hat{s}_{\text{UN}}] = \frac{\sum_i (1 - \pi_i^+ + \pi_i^-)^2 \pi_i^+ (\pi_i^- + m_0 + n_0 - n_1 \pi_i^- - n_1 \pi_i^+)}{n_1 (n_1 + m_1)(\pi_i^+ + \pi_i^-)^2 (1 - \pi_i^+ + \pi_i^-)}
\]

(11)

\[
\text{var}[\hat{s}_{\text{EU}}] = \frac{\sum_i (1 - \pi_i^+ + \pi_i^-)^2 \pi_i^+ (\pi_i^- + m_0 + n_0 - n_1 \pi_i^- - n_1 \pi_i^+)}{n_1 (n_1 + m_1)(\pi_i^+ + \pi_i^-)^2 (1 - \pi_i^+ + \pi_i^-)}
\]

(12)

For the estimators of \(\phi_E^{+\text{UN}}\) and \(\phi_E^{-\text{UN}}\)

\[
\text{var}[\hat{\phi}_E^{+\text{UN}}] = \frac{\sum_i \pi_i^+ (m_1 + \pi_i^- + \pi_i^+)(\pi_i^- + m_0 + n_0 - n_1 \pi_i^- - n_1 \pi_i^+)}{n_1 (n_1 + m_1)(\pi_i^+ + \pi_i^-)^2 (1 - \pi_i^+ + \pi_i^-)}
\]

(13)

\[
\text{var}[\hat{\phi}_E^{-\text{UN}}] = \frac{\sum_i \pi_i^+ (m_1 + \pi_i^- + \pi_i^+)(\pi_i^- + m_0 + n_0 - n_1 \pi_i^- - n_1 \pi_i^+)}{n_1 (n_1 + m_1)(\pi_i^+ + \pi_i^-)^2 (1 - \pi_i^+ + \pi_i^-)}
\]

(14)

If no baseline data are available, the MLEs and expressions for the asymptotic variances can be obtained by substituting \(m_1 = m_0 = 0\) in Equation (10) and Equations (11-14), respectively. We note here that even when it is available, the MLE for \(\phi_E^{+\text{UN}}\) and the expression for its asymptotic variance given by Equation (14) do not depend on the baseline data. However, the asymptotic variances of, \(s_{\text{UN}}\), \(s_{\text{EU}}\), \(\phi_E^{+\text{UN}}\), and \(\phi_E^{-\text{UN}}\) are always reduced by including the baseline data.

Equations (11-14) are used to derive the standard errors for the estimates of sensitivity and specificity of the believe the positive protocol, and for the log-ratios of the positive and negative likelihood ratios of the two tests, by replacing the values of \(\pi_i^+\), and \(\pi_i^-\), \(D=0,1\), with their corresponding MLEs, and then taking the square root. In cases where the sample size for the assessment study is small or the counts for some \(n_i^+\) are very small, other methods such as bootstrapping are recommended for obtaining the standard errors [7].

**Believe the negative protocol**

Believe the negative protocols are used to reduce the number of false
positives and therefore increases the specificity over the existing test. We will denote the protocol as $E \cap N$. The conditional probabilities, given the true disease status, of the existing test and the believe the negative protocol to improve sensitivity over the use of the existing test alone. In practice the protocols can be implemented using either an add-on or triage plan. In an add-on plan the existing test is first given to all subjects. Follow-up testing with the new test is conducted only on those with an initial negative result for believe the positive or initial positive result for believe the negative. For a triage plan, the new test is used first and the results determine who will receive the follow-up testing with the new test. Depending on the choice of test and protocol combination depends on the basis of a positive result from the existing test alone. This combination means that only subjects likely to be truly diseased (on the basis of a positive result from the existing test) are given the new test. The diagnostic properties of the protocols do not depend on whether they will be used with an add-on or triage plan.

Case-control versus cohort studies: The expressions for the MLEs in Equations (10) and (20) and the asymptotic variances in Equations (11-14) and Equations (21-24) apply to settings where the numbers of diseased, $n_D$, and non-diseased, $n_N$, in the assessment and baseline studies are fixed, as in a case-control design. If the assessment study is designed as a cohort study, these expressions still apply, with $m_D = m_N = 0$. For a triage plan, the new test is used first and the results determine who will receive the follow-up testing with the new test. Depending on the choice of test and protocol combination depends on the basis of a positive result from the existing test alone. This combination means that only subjects likely to be truly diseased (on the basis of a positive result from the existing test) are given the new test. The diagnostic properties of the protocols do not depend on whether they will be used with an add-on or triage plan.

Paired versus sequential assessment studies: The analysis methodology given above assumes that a paired assessment study is used. If it is known a priori whether the new and existing tests will be combined using a believe the positive or believe the negative protocol a sequential assessment study can be used instead. We still assume full verification via a gold standard. First, the existing test should be applied to all subjects. This is necessary so that we can test for equivalence of the diagnostic properties of the existing test across the baseline data and assessment study. Following that, the new test should be applied to only subjects in the assessment study and the baseline data, the expressions for the asymptotic variance for the estimations of $s_{\text{SENS}}$, $s_{\text{SPEC}}$, $\phi_{E/N}$, and $\phi_{E \cap N}$ are

$$
\text{var}[s_{\text{SENS}}] = \pi_1^2 \left[ \frac{1}{n_D} \left( \frac{1}{n_D} + \frac{1}{n_N} \right) \right] \left[ \frac{1}{n_D} \left( \frac{1}{n_D} + \frac{1}{n_N} \right) \right] \left[ \frac{1}{n_D} \left( \frac{1}{n_D} + \frac{1}{n_N} \right) \right]
$$

$$
\text{var}[s_{\text{SPEC}}] = \pi_1^2 \left[ \frac{1}{n_D} \left( \frac{1}{n_D} + \frac{1}{n_N} \right) \right] \left[ \frac{1}{n_D} \left( \frac{1}{n_D} + \frac{1}{n_N} \right) \right] \left[ \frac{1}{n_D} \left( \frac{1}{n_D} + \frac{1}{n_N} \right) \right]
$$

$$
\text{var}[\phi_{E/N}] = \left[ \frac{1}{n_D} \left( \frac{1}{n_D} + \frac{1}{n_N} \right) \right] \left[ \frac{1}{n_D} \left( \frac{1}{n_D} + \frac{1}{n_N} \right) \right] \left[ \frac{1}{n_D} \left( \frac{1}{n_D} + \frac{1}{n_N} \right) \right]
$$

$$
\text{var}[\phi_{E \cap N}] = \left[ \frac{1}{n_D} \left( \frac{1}{n_D} + \frac{1}{n_N} \right) \right] \left[ \frac{1}{n_D} \left( \frac{1}{n_D} + \frac{1}{n_N} \right) \right] \left[ \frac{1}{n_D} \left( \frac{1}{n_D} + \frac{1}{n_N} \right) \right]
$$

There is a clear symmetry between these results and those for the believe the positive protocol in Equations (11-14). If baseline data are not available, the MLEs can be obtained by substituting $m_D = m_N = 0$ into the relevant equations. For the believe the negative protocol the MLE for $\phi_{E \cap N}$ and the expression for its asymptotic variance given by Equation (23) do not depend on the baseline data.

Additional Considerations

Add-on and triage plans: The believe the positive and believe the negative protocols determine the rule for combining the results of the two tests. In practice the protocols can be implemented using either an add-on or triage plan. In an add-on plan the existing test is first given to all subjects. Follow-up testing with the new test is conducted only on those with an initial negative result for believe the positive or initial positive result for believe the negative. For a triage plan, the new test is used first and the results determine who will receive the follow-up testing with the new test. Depending on the choice of test and protocol combination depends on the basis of a positive result from the existing test alone. This combination means that only subjects likely to be truly diseased (on the basis of a positive result from the existing test) are given the new test. The diagnostic properties of the protocols do not depend on whether they will be used with an add-on or triage plan.

Case-control versus cohort studies: The expressions for the MLEs in Equations (10) and (20) and the asymptotic variances in Equations (11-14) and Equations (21-24) apply to settings where the numbers of diseased, $n_D$, and non-diseased, $n_N$, in the assessment and baseline studies are fixed, as in a case-control design. If the assessment study is designed as a cohort study, these expressions still apply, with $n_D$ replaced by $n_D \theta$ and $n_N$ by $n_N (1 - \theta)$, where $\theta$ is the prevalence of disease and $n$ is the total sample size. Similar adjustments can be made to $m_D$ and $m_N$ if the baseline data come from a cohort study.

Paired versus sequential assessment studies: The analysis methodology given above assumes that a paired assessment study is used. If it is known a priori whether the new and existing tests will be combined using a believe the positive or believe the negative protocol a sequential assessment study can be used instead. We still assume full verification via a gold standard. First, the existing test should be applied to all subjects. This is necessary so that we can test for equivalence of the diagnostic properties of the existing test across the baseline data and assessment study. Following that, the new test should be applied to only subjects in the assessment study and the baseline data, the expressions for the asymptotic variance for the estimations of $s_{\text{SENS}}$, $s_{\text{SPEC}}$, $\phi_{E/N}$, and $\phi_{E \cap N}$ are

$$
\text{var}[s_{\text{SENS}}] = \pi_1^2 \left[ \frac{1}{n_D} \left( \frac{1}{n_D} + \frac{1}{n_N} \right) \right] \left[ \frac{1}{n_D} \left( \frac{1}{n_D} + \frac{1}{n_N} \right) \right] \left[ \frac{1}{n_D} \left( \frac{1}{n_D} + \frac{1}{n_N} \right) \right]
$$

$$
\text{var}[s_{\text{SPEC}}] = \pi_1^2 \left[ \frac{1}{n_D} \left( \frac{1}{n_D} + \frac{1}{n_N} \right) \right] \left[ \frac{1}{n_D} \left( \frac{1}{n_D} + \frac{1}{n_N} \right) \right] \left[ \frac{1}{n_D} \left( \frac{1}{n_D} + \frac{1}{n_N} \right) \right]
$$

$$
\text{var}[\phi_{E/N}] = \left[ \frac{1}{n_D} \left( \frac{1}{n_D} + \frac{1}{n_N} \right) \right] \left[ \frac{1}{n_D} \left( \frac{1}{n_D} + \frac{1}{n_N} \right) \right] \left[ \frac{1}{n_D} \left( \frac{1}{n_D} + \frac{1}{n_N} \right) \right]
$$

$$
\text{var}[\phi_{E \cap N}] = \left[ \frac{1}{n_D} \left( \frac{1}{n_D} + \frac{1}{n_N} \right) \right] \left[ \frac{1}{n_D} \left( \frac{1}{n_D} + \frac{1}{n_N} \right) \right] \left[ \frac{1}{n_D} \left( \frac{1}{n_D} + \frac{1}{n_N} \right) \right]
$$

There is a clear symmetry between these results and those for the believe the positive protocol in Equations (11-14). If baseline data are not available, the MLEs can be obtained by substituting $m_D = m_N = 0$ into the relevant equations. For the believe the negative protocol the MLE for $\phi_{E \cap N}$ and the expression for its asymptotic variance given by Equation (23) do not depend on the baseline data.
those subjects with an initial negative result for a believe the positive protocol or initial positive result for a believe the negative protocol. This means we do not observed the full 2x2 data tables. However, the MLEs and asymptotic variance expressions under a believe the positive protocol in Equations (10-14) only depend on $n_{10}^0$, $n_{01}^0$ and $n_0$, so it is not necessary to separately observe $(n_{10}^0, n_{01}^0)$, $D=1.0$. The same holds for a believe the negative protocol where it is not necessary to separately observe $(n_{01}^0, n_{10}^0)$, $D=1.0$.

Application

As an example, we consider breast cancer detection in symptomatic women. In a case-control study, Houssami et al. [16] examined the mammography (Mx) and sonography/ultrasound (US) test results of 240 women shown to have breast cancer and 233 age-matched controls. Both cases and controls were women aged 25 to 55 years who had been referred to testing due to the presence of symptoms (a palpable lump, pain or localized discomfort, etc.). The controls were selected among those who were not diagnosed with breast cancer in the two years following their assessment. The images were interpreted by two radiologists blinded to the subject’s cancer status. Since both the cases and controls were tested using both modalities, the paired data are available in Table 1. A complete discussion of the study design and inclusion and exclusion criteria is available in Houssami et al. [16]. While the original purpose of the study was to investigate the age-specific sensitivity and specificity of the tests, we will use the data as an illustration to evaluate whether ultrasound is useful when combined with mammography in a believe the positive or believe the negative protocol. Based on the paired data in Table 1, $\hat{\text{sens}}_{\text{US}}=75.8\%$, $\hat{\text{spec}}_{\text{US}}=87.6\%$, $\hat{\text{sens}}_{\text{Mx}}=81.6\%$, and $\hat{\text{spec}}_{\text{Mx}}=88.0\%$. To illustrate, we treat mammography as the existing test and ultrasound as a new test. Among the cases and controls, 34.1% and 17.6%, respectively, of the women had discrepant mammography and ultrasound test results.

In order to proceed with the methods described in the previous section, we need baseline data on the performance of mammography in the given population. We use the data provided in Kavanagh et al. [17] for this purpose (see Table 1-Baseline data) which provide estimates of the sensitivity and specificity of mammography in a similar symptomatic population. We consider the subpopulation of women with any breast symptoms and estimate the sensitivity and specificity of mammography to be 71.0% and 89.7%, respectively. Both studies were conducted among Australian women in the mid 1990’s. However, the population in the baseline study was slightly older which may cause an inflation of the sensitivity estimate. However, the likelihood ratio test derived in Appendix A of the Supplementary Materials gave no evidence ($p$-value=0.35) to reject the hypothesis that the two accuracy measures are common for the assessment study and baseline population. We consider two diagnostic protocols:

1. Mx $\cup$ US, a believe the positive protocol with US as an add-on to Mx. This protocol will have better sensitivity than Mx alone and uses US to catch potential initial false negatives.

2. US $\cap$ Mx, a believe the negative protocol with US as a triage for Mx. The protocol will have better specificity than Mx alone and in practice would greatly reduce the number of mammograms used in a screening program.

The MLEs for the sensitivity and specificity of these diagnostic protocols under both a standard analysis (without baseline data) and an augmented analysis (with the inclusion of baseline data) are shown in Table 2. Here, we are interested in assessing the gain in precision in estimating the sensitivity and specificity of the new protocols when we include the baseline data in the analysis. We note that, for the believe the positive protocol, the estimates of both sensitivity and specificity are similar with and without the inclusion of the baseline data (around 96% sensitivity and 80% specificity). Also, the precision of the estimator of sensitivity does not change noticeably when we include the baseline data. However, the standard error for the estimate of the specificity decreases by 30%. For the believe the negative protocol, the estimates given by the two analyses are also similar. There is a decrease of 10% in the standard error of the sensitivity estimate and 25% for the specificity estimate when the baseline data are included in the analysis.

For both protocols, the estimates of the logarithm of the ratio of positive and negative likelihood ratios and their associated standard errors for the standard and augmented analysis are also given in Table 2. For the believe the positive protocol, the MLE for the logarithm of the ratio of positive likelihood ratios, $\hat{f}_{Mx,US}$, is negative for both the standard and augmented analyses, although it is more extreme for the augmented analysis (i.e., -0.290 versus -0.359). The standard error of the estimate of $\hat{f}_{Mx,US}$ decreases by around 17% when the baseline data are used in estimation. The estimate of the log-ratio of the negative likelihood ratios for the believe the positive protocol, $\hat{f}_{Mx,US}$, is identical for the standard and augmented analyses (i.e., -1.655). The corresponding standard errors are very similar, with a slight decrease when the baseline data are used (about 4%).

When ultrasound is included in a believe the negative protocol, the MLE of the log-ratio of the negative likelihood ratios, $\hat{f}_{US,Mx}$, given by the augmented analysis is smaller than the one given by the standard analysis (0.347 compared to 0.363). The standard error of the estimate decreases by 17% when the baseline data are used. The estimates of the log-ratio of the positive likelihood ratios, $\hat{f}_{US,Mx}$, are identical (i.e. 1.081) for the standard and augmented analyses. However, the corresponding asymptotic standard error is slightly larger for the augmented analysis than for the standard analysis. To investigate this counterintuitive result, we conducted a simulation study in which the true parameter values and sample sizes are assumed to equal the relevant proportions from the three 2x2 tables in Table 1. The results, given in Table 3, indicate that the asymptotic standard error from the standard analysis underestimates the standard deviation corresponding to $\hat{f}_{US,Mx}$, (0.303 versus 0.342). Recall that the expressions for $\hat{f}_{US,Mx}$, (also $\hat{f}_{US,Mx}$) and its asymptotic standard deviation do not depend on the baseline data. We note a similar situation for the standard error of the estimate of $\hat{f}_{Mx,US}$ from both the augmented and standard analysis.

<table>
<thead>
<tr>
<th>Sensitivity</th>
<th>Specitivity</th>
<th>$\hat{f}_{Mx,US}$</th>
<th>$\hat{f}_{US,Mx}$</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sensitivity</td>
<td>Specitivity</td>
<td>$\hat{f}_{Mx,US}$</td>
<td>$\hat{f}_{US,Mx}$</td>
</tr>
<tr>
<td>Standard Analysis</td>
<td>0.958 (0.013)</td>
<td>0.790 (0.027)</td>
<td>-0.290 (0.125)</td>
</tr>
<tr>
<td>Augmented Analysis</td>
<td>0.955 (0.013)</td>
<td>0.808 (0.019)</td>
<td>-0.359 (0.102)</td>
</tr>
</tbody>
</table>

Table 2: Estimated sensitivity, specificity, and log-ratio of likelihood ratios ($\hat{f}$ and $\hat{f}$) and (standard errors) for mammography (Mx) versus believe the positive (Mx US) and believe the negative (US Mx) protocols.
which underestimates the corresponding standard deviation (0.289 versus 0.317 for the standard analysis). The simulation study also suggests that the other asymptotic standard errors reported in Table 2 are good approximations to their corresponding standard deviations.

**Simulation Study**

In the previous section, for the breast cancer screening example, we found that including baseline information generally improved the precision of the estimators of sensitivity and specificity of the new protocols, and of the logarithm of the ratio of positive likelihood ratios (for believe the positive) and negative likelihood ratios (for believe the negative). In order to investigate the magnitude of the potential precision gains across a variety of baseline and assessment study sizes and diagnostic accuracies, we conducted a factorial simulation study.

We considered four study sizes for the baseline data $m_0, m_1=100, 250, 1000, 5000$ and two sizes for the paired assessment study $n_0, n_1=100, 250$. Diagnostic accuracies of 70%, 80%, 90% were considered for each of $\text{sens}_E$, $\text{spec}_E$, $\text{sens}_N$ and $\text{spec}_N$. In addition, the level of agreement between the two tests, represented through $\pi_0 = \Pr\{E=1, N=1\} = \Pr\{D=1\}$ and $\pi_1 = \Pr\{E=0, N=0\}$ were set at 10%, 50%, and 90% of the way between their minimum and maximum values given by (25) and (26):

\[
\max(0, \pi_0 + \pi_1 - 1, 0) \leq \Pr\{E=1, N=1\} = \Pr\{D=1\} \leq \min(1, \pi_0 + \pi_1) \quad (25)
\]

\[
\max(\pi_1, \pi_0 - 1, 0) \leq \Pr\{E=0, N=0\} = \Pr\{D=0\} \leq \min(\pi_1, \pi_0) \quad (26)
\]

This leads to a total of 46,656 scenarios. In each scenario the ratio of the asymptotic standard deviation of the estimators of the sensitivity, specificity, $\phi^+$ and $\phi^-$ for an augmented (with baseline data) versus standard (no baseline data) analysis were calculated. Results showing the ratios of asymptotic standard deviations for a believe the positive protocol are presented in Figure 1 for varying baseline sample sizes $m_0$ and $m_1$. Results are similar for a believe the negative protocol with increased precision seen for $\phi^+$ rather than $\phi^-$. Over the scenarios considered, the median reductions in asymptotic standard deviation for the sensitivity, specificity, and $\phi^+$ were 11%, 19%, and 19%, respectively.

The greatest precision improvements are found with larger baseline study sizes with the number of diseased $m_1$ impacting the precision of $\text{sens}_{E,N}^-$. The number of non-diseased impacting the precision of $\text{spec}_{E,N}^-$. The gains in precision in the estimation of $\text{sens}_{E,N}^+$ also tend to be higher when the number of diseased subjects in the assessment study $n_1$ is small, $\text{sens}_N$ is high, $\text{spec}_N$ is low, and the level of agreement between $\text{sens}_E$ and $\text{spec}_E$ is high. Conversely, the gains in precision in the estimation of $\text{sens}_{E,N}^-$ are greatest when $\text{spec}_N$ is high, $\text{spec}_E$ is low, and their agreement is low.

**Discussion**

In this paper, we provided methods for incorporating available baseline information into the estimation of the sensitivity and specificity of a new believe the positive or believe the negative protocol, and of the logarithm of the ratios of positive and negative likelihood ratios of the new protocol vs. the existing test. Comparison between the likelihood ratios of the new protocol and the existing test is equivalent to a comparison between the corresponding predictive values when applied to the same population. Asymptotic variances were derived for all proposed statistics.

Throughout the paper, we assumed that baseline data for the augmented analysis were available in the form of a 2×2 table. Alternatively, baseline data might only be available in the form of estimated sensitivity and specificity and their standard errors or confidence intervals. Provided the number of subjects is given, it is straightforward to transform a test's sensitivity and standard error to the data $m_0^+, d=0$ using formulas for the MLE and variance of a proportion from a binomial distribution. Additionally, the methods presented here could be expanded to include a second set of baseline data perhaps from a preliminary evaluation of the new test.

Before conducting an augmented analysis we propose using a likelihood ratio test of the equivalence of the diagnostic properties of the existing test across the baseline and assessment study data. If this test is rejected it suggests that the existing test is behaving differently between subjects in the baseline and assessment studies. This could be due to inherent differences in the populations (for example age, disease strain, disease severity) or differences in the application of the test. In this case it is unwise to incorporate the baseline data in the analysis and a standard analysis should be used by substituting $m_1=m_2=0$ into the formulas given in the Methods section.

Incorporating baseline data increases the precision of the estimators of the sensitivity and specificity of the protocol and of the log-ratio of negative (positive) likelihood ratios of the believe the positive (negative) protocol versus the existing test. We do not have an intuitive explanation as to why the MLE and asymptotic variance of $\phi^+_{E,N}$ and $\phi^-_{E,N}$ do not depend on the baseline data. However, in a believe the positive protocol, we get higher sensitivity due to an increased number of positive results, we may be more interested in comparing the positive predictive values of $E\cup N$ and the existing test through $\phi^+_{E,N}$ whose estimation and precision does benefit from the inclusion of baseline data.

In terms of planning an assessment study, there are two key ways in which incorporating available information is beneficial. First, for

<table>
<thead>
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<th>$\phi^-$</th>
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<tbody>
<tr>
<td>Est</td>
<td>ESE</td>
<td>ASE</td>
<td>Est</td>
</tr>
<tr>
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<td>0.013</td>
<td>0.013</td>
</tr>
<tr>
<td>Augmented Analysis</td>
<td>0.955</td>
<td>0.014</td>
<td>0.013</td>
</tr>
</tbody>
</table>

Table 3: Estimated sensitivity, specificity and log-ratio of likelihood ratios ($\phi^+$ and $\phi^-$), empirical standard errors (ESE) and asymptotic standard errors (ASE) for mammography (Mx) versus believe the positive (Mx U US) and believe the negative (US \(\cap\) Mx) protocols from 50,000 simulations based on parameter values derived from the breast cancer example.
a fixed assessment study size, using baseline data will improve the precision of the estimators of important quantities for the combined protocol. Second, for a fixed desired precision, the use of baseline data may substantially decrease the required sample sizes for the assessment study. Initial estimates of the properties of the new and existing test along with the expressions for asymptotic variances given in the Methods section can be used for planning purposes.

If the assessment study is run using a paired design the properties of all four combinations of protocol (believe the positive or believe the negative) and plan (add-on or triage) can be estimated. If the protocol is fixed a priori a sequential assessment study can be run with the existing test first applied to all subjects. This study mimics the use of an add-on plan and gives all the necessary information to compare the existing test with the new protocol, for both augmented and standard analysis. The results apply to either plan. However, in a sequential study mimicking a triage plan only the subjects testing positive with the new test are further tested with the existing test. Therefore, the sensitivity and specificity of the existing test cannot be estimated from the assessment study and therefore cannot be compared to the baseline results. An augmented analysis could still be conducted using methods similar to those developed in the Methods section.

In the example with breast cancer detection, neither the existing test nor the believe the positive protocol outperformed the other in terms of both positive and negative predictive values. In such cases
Macaskill et al. [7] propose choosing between the existing test and the protocol based on the expected numbers of additional false positive and true positive results identified by the believe the positive protocol. As this protocol involves testing with the new test the subjects who tested negative for the existing test, the believe the positive protocol will generate some additional false positives and true positives when compared to the results of the existing test only. For a believe the negative protocol one would compare the expected numbers of additional false negative and true negative results identified by the protocol. We do not further investigate this trade-off here but it may warrant future attention.

**Supplementary Materials**

Appendix A gives the derivation of a likelihood ratio test for testing the equivalence in the diagnostic properties (sensitivity and specificity) of the existing test across the baseline data and assessment study. Appendix B gives the derivation of the asymptotic variances of the estimators of the sensitivity, specificity and log-ratios of the positive and negative likelihood ratios for a believe the positive protocol.

**Acknowledgments**

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**References**