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# 1 Targeted Nanoparticle Binding & Detection in Petroleum Hydrocarbon 2 Impacted Porous Media

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## 9 Abstract

10 Targeted nanoparticle binding has become a core feature of experimental pharmaceutical product  
11 design which enables more efficient payload delivery and enhances medical imaging by accumulating  
12 nanoparticles in specific tissues. Environmental remediation and geophysical monitoring encounter  
13 similar challenges which may be addressed in part by the adoption of targeted nanoparticle binding  
14 strategies. This study illustrates that engineered nanoparticles can bind to crude oil-impacted silica sand,  
15 a selective adsorption driven by active targeting based on an amphiphilic polymer coating. This coating  
16 strategy resulted in 2 mg/kg attachment to clean silica sand compared to 8 mg/kg attachment to oil-  
17 impacted silica sand. It was also shown that modifying the surface coating influenced the binding  
18 behaviour of the engineered nanoparticles – more hydrophobic polymers resulted in increased binding.  
19 Successful targeting of Pluronic-coated iron oxide nanoparticles to a crude oil and silica sand mixture  
20 was demonstrated through a combined quantitative Orbital Emission Spectroscopy mass analysis  
21 supported by Vibrating Scanning Magnetometer magnetometry, and a qualitative X-ray micro-computed  
22 tomography (CT) visualization approach. These non-destructive characterization techniques facilitated  
23 efficient analysis of nanoparticles in porous medium samples with minimal sample preparation, and in  
24 the case of X-Ray CT, illustrated how targeted nanoparticle binding may be used to produce 3-D images  
25 of contaminated porous media. This work demonstrated successful implementation of nanoparticle

26 targeted binding toward viscous LNAPL such as crude oil in the presence of a porous medium, a step  
27 which opens the door to successful application of targeted delivery technology in environmental  
28 remediation and monitoring.

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## 29 **Introduction**

30 Targeted binding is a familiar concept in the field of healthcare which typically refers to specific analytes  
31 preferentially binding to specific receptors on cells and tissues(Davis et al., 2008). This observation has  
32 driven innovation in nanomedicine over the past two decades and resulted in functionalized  
33 nanoparticles specifically modified to target diseased tissues and release therapeutic payloads(Petros  
34 and DeSimone, 2010; Srinivasarao et al., 2015). Implementation of a targeted nanoparticle binding  
35 platform provides direct enhancement to therapeutic efficiency and reduces side effects by minimizing  
36 drug action on healthy tissue. Targeted binding can also drive accumulation of nanoparticles in tissue  
37 with specific physical or chemical characteristics which can then be detected using medical imaging  
38 technologies and provide an image of target areas(Gupta and Gupta, 2005; Qin et al., 2007; Rosen et al.,  
39 2012). Soil contamination and its remediation bears many similar challenges to diseased tissue and its  
40 treatment; the impacted site is often not easily accessible, *in situ* treatment wishes to avoid delivering a  
41 payload to clean soil, and the contaminated region has different chemical and physical characteristics  
42 from the surrounding material. By taking advantage of the unique physical and chemical properties of  
43 the impacted region, nanoparticles can be designed to preferentially accumulate there, allowing for  
44 targeted treatment or enhanced imaging.

45 Soil remediation using materials designed for targeted removal of specific contaminants has been a  
46 subject well studied in the area of toxic metals removal wherein the mobile contaminant species is  
47 adsorbed to a high-surface area nanostructure such as titanate nanotubes or modified graphene  
48 oxide(Chang et al., 2017a; Hu et al., 2017a, 2017c; Linghu et al., 2017). Some graphene oxide-based  
49 strategies introduce the possibility of target contaminant immobilization by forming graphene oxide-  
50 metal ion complexes with enhanced adsorption to mineral material(Chang et al., 2017b; Hu et al.,  
51 2017b; Sheng et al., 2018). Treatment strategies pursuing elimination of fixed contaminants, such as in  
52 the case of viscous non-aqueous phase liquids (NAPLs), require the treatment agent to be emplaced in

53 the contaminated region rather than adsorb the contaminant to its surface, and a knowledge gap exists  
54 in this regard. Soil contamination by NAPL petroleum hydrocarbons (PHCs) often requires some type of  
55 remedial activity to address potential impacts on ecosystems or human health. While conventional *in*  
56 *situ* remediation technologies utilizing direct delivery of reagents have been successful in treating some  
57 types of PHCs (Jawitz et al., 1998; Salanitro et al., 2000; Sra et al., 2013), options are limited for viscous  
58 hydrocarbons such as heavy crude oils, for which no effective *in situ* treatment strategy exists.

59 Advancements in nanotechnology have led to the emergence of nanoremediation which utilizes the  
60 higher specific surface area of nanoparticles to achieve faster treatment, but applications are still limited  
61 by other drawbacks such as particle self-aggregation and aggregation with mineral material (Karn et al.,  
62 2009; Kuppusamy et al., 2016). Nanoremediation treatment of a contaminant may occur directly as a  
63 result of nanoparticle contact, as in the case of nZVI (Pardo et al., 2015), or indirectly through Fenton-like  
64 catalysis of a persulfate reaction where the nanoparticle supplies leached iron ions (Al-Shamsi and  
65 Thomson, 2013). The challenge of delivering nanoparticles to a treatment zone has spurred significant  
66 investigation of coating materials to improve their transport properties (Braun et al., 2015; Johnson et  
67 al., 2013; Li et al., 2015; Luna et al., 2015; Su et al., 2015); however, these improvements do not  
68 guarantee nanoparticle delivery directly to the NAPL interface, and have led to serious discussion  
69 regarding the toxicity risks of releasing mobile nanoparticles into an unrestricted environment (Höss et  
70 al., 2015; Karn et al., 2009; Tosco et al., 2014). Based on similarities to nanomedicine, *in situ*  
71 nanoremediation may be improved by the adoption of targeted binding. To this end, a successful  
72 targeted nanoparticle delivery system must be capable of: (1) using nanoparticles that can be tuned to  
73 stick or bind to target NAPL mass that they contact, (2) transporting functional nanoparticles in various  
74 geologic media over significant distances (i.e., > 1 m), and (3) allowing the bound nanoparticles to be  
75 triggered either actively or passively to enable treatment.

76 In general, targeted delivery is divided into two broad categories: passive and active which rely on  
77 physical and chemical surface properties of the target zone, respectively(Bertrand et al., 2014; Elkhodiry  
78 et al., 2016; Ganta et al., 2008). The targeting mechanism for NAPL PHCs presented in this paper is  
79 similar to active targeting seen in drug delivery and achieved by tailoring the nanoparticle surface to  
80 interact with the NAPL PHC interface(Saleh et al., 2006). Such an 'active targeting' technique has been  
81 previously demonstrated for chlorinated hydrocarbon NAPLs using a block co-polymer nanoparticle  
82 surface coating capable of stabilizing nanoparticles in aqueous suspension and promoting hydrophobic  
83 interactions(Saleh et al., 2006, 2005; Wang and Acosta, 2013). Targeted delivery of nanoparticles has  
84 the potential to further increase the impact and efficiency of nanoremediation techniques by  
85 concentrating the reagent in direct contact with the NAPL.

86 In this paper, we report on the selective binding of functional nanoparticles to one class of NAPL PHCs.  
87 Specifically, we focussed on the ability of an inexpensive, non-toxic(Singh-Joy and McLain, 2016),  
88 amphiphilic block co-polymer coating (Pluronic) applied to iron oxide nanoparticles to promote aqueous  
89 stability and hydrophobic interactions with viscous hydrocarbons. Iron oxide nanoparticles were  
90 selected as the core material due to their simple synthesis, possible application as an iron catalyst for  
91 various chemical oxidation processes (Usman et al., 2012), and capability to act as an energy absorber  
92 for thermal treatment strategies (Chien, 2012; Kharisov et al., 2012; Maehara et al., 2005; Zhou et al.,  
93 2016). Nanoparticle targeted binding behaviour was assessed using a batch experimental design and  
94 quantified indirectly using aqueous-phase concentration measurements. This indirect approach was  
95 supported by direct quantification of bound nanoparticles using VSM magnetometry, and binding was  
96 further visualized using a qualitative X-ray Computed Tomography (CT) technique.

97 Nanotechnology-enabled sensing techniques are a subject of significant research for detecting  
98 contaminants. Through interaction of the nanoparticle with an analyte of interest, a detectable signal  
99 can be generated to confirm or measure the presence of the analyte(Zeng et al., 2016; Zhu et al., 2014).

100 Using nanoparticles adsorbed to target contaminants in porous media as a contrast agent is a nano-  
101 enabled sensing technique analogous to X-Ray CT medical imaging. In the context of geosciences, X-Ray  
102 CT relies on identifying areas of varying radiological density which can vary with mineral type (Cnudde et  
103 al., 2006), for example,  $\text{Fe}_3\text{O}_4$  absorbs X-Rays more strongly than  $\text{SiO}_2$ . The possibility and threshold of  
104 detection for nanoparticles in porous media is a key question that must be addressed in the context of  
105 X-Ray CT detection of nanoparticles in geologic media. Typical characterization of nanoparticles or trace  
106 metals embedded in soil relies on tedious, destructive techniques such as acid digestion and subsequent  
107 ICP-MS or ICP-OES analysis (Liu et al., 2017, 2018), prompting research into new analysis techniques  
108 which avoid lengthy sample preparation (Kaveh and Beauchemin, 2014; Russo et al., 2002; Senesi et al.,  
109 2009). 3-D mapping of nanoparticle accumulation would require performing this type of lengthy analysis  
110 multiple times in order to construct a model. With the X-Ray CT technique introduced here, non-  
111 destructive 3-D analysis of relatively large samples is achieved with a single, quick, high-resolution scan  
112 and minimal sample preparation. This research effort investigates one of the critical features of a  
113 potential targeted nanoparticle delivery system, i.e., ability to bind to a target NAPL mass, and  
114 demonstrates what a possible end-use of nanoparticle targeted binding might look like through the  
115 application of X-Ray CT to detect contaminated sands containing bound nanoparticles. Research that  
116 addresses the other two features of a robust targeted delivery platform (transport in various porous  
117 media, and activation options for treatment) is ongoing.

118

## 119 **Materials and Methods**

### 120 **Materials**

121 Iron (II) sulfate heptahydrate ( $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$ , > 99%), iron (III) chloride hexahydrate ( $\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$ , > 99%),  
122 ammonium hydroxide ( $\text{NH}_4\text{OH}$ , 28-30% in water), dichloromethane (DCM, > 99%), and oleic acid (> 90%)  
123 were purchased from Sigma Aldrich (St. Louis, MO, USA). Hydrochloric acid (37%) and ethanol (ACS

124 grade, 99%) were purchased from Fisher Scientific (Hampton, NH, USA). Pluronic co-polymers P104, L62,  
125 and L121 (Table 1) were gifted by Brenntag (Essen, Germany) and BASF (Ludwigshafen, Germany). Silica  
126 sand (99.7% SiO<sub>2</sub>, d<sub>50</sub> = 0.2 mm) was purchased from Opta Minerals, Inc. (Hamilton, ON, Canada). Crude  
127 oil (heavy, sour crude, API gravity 13.3 ( $\rho = 0.977 \text{ g/cm}^3$ ), kinematic viscosity  $4.745 \times 10^{-4} \text{ m}^2/\text{s}$  @ 40 °C)  
128 was provided by Chevron Energy Technology Company (Houston, TX, USA). All materials were used as  
129 received.

### 130 Nanoparticle Synthesis

131 Nanoparticle synthesis was adapted from several methods previously reported (Chan and Gu, 2013; Cui  
132 et al., 2007; Xu et al., 2006). FeSO<sub>4</sub>·7H<sub>2</sub>O and FeCl<sub>3</sub>·6H<sub>2</sub>O were added to deoxygenated water at a molar  
133 ratio of 2:3 (FeSO<sub>4</sub>:FeCl<sub>3</sub>). Sufficient NH<sub>4</sub>OH and oleic acid were added to achieve final concentrations of  
134 4 and 0.22 mol/L, respectively. This solution was stirred at 70 °C for 1 h, then stirred at 90 °C under  
135 flowing N<sub>2</sub> for 1 h to purge evolved NH<sub>3</sub> gas. After cooling to room temperature, the black, magnetic  
136 precipitate was recovered by magnetic decantation and washed 3x by deoxygenated Millipore DI water  
137 (Millipore Elix 5), and then 3x by ethanol before being dried under flowing N<sub>2</sub>.

### 138 Nanoparticle Phase Transfer

139 A mixture of oleic acid in hexane (1% v/v) was prepared to which dried nanoparticles were added at a  
140 concentration of 90 g/L and sonicated for 10 min (VWR "Symphony" 1.9 L Ultrasonic Cleaner). The  
141 resulting suspension was added to various aqueous solutions of Pluronic co-polymer(s) at a  
142 concentration of 10 % v/v and probe sonicated for 30 min to emulsify the water and hexane phases  
143 (Fisher Scientific FB505 Sonic Dismembrator, 200 W). The emulsified nanoparticle sample was  
144 transferred to a separatory funnel and separated over 48 h. The resulting aqueous nanoparticle  
145 suspension (~3 g/L) was stored until used.

### 146 Nanoparticle Binding Studies

147 Oil-impacted silica sand (OSS) was prepared by dissolving crude oil in DCM at a concentration of 60 g/L,  
148 mixing it with dry silica sand, and evaporating the DCM such that the final concentration of crude oil in



149 sand was 1.5% (m/m). 15 g of OSS was then added to a 20-mL cylindrical glass vial, followed by 10 mL of  
150 nanoparticle suspension diluted with Millipore DI water to either 1:100 or 1:10 of its concentration after  
151 phase-transfer. The glass vials were then loaded onto an orbital shaker (Stovall Life Sciences “Belly  
152 Dancer”) for 48 h. Following mixing, the nanoparticle solution was recovered by filtration (1.5  $\mu\text{m}$  glass  
153 fiber, Whatman). The retentate was washed with 100 mL Millipore DI water and dried by vacuum  
154 desiccation for 96 h before CT, scanning electron microscopy (SEM)/energy dispersive X-ray  
155 spectroscopy (EDS), or Vibrating Scanning Magnetometer/ Superconducting Quantum Interference  
156 Device (VSM/SQUID) analysis. For control, a “clean” silica sand (SS) sample was prepared identically as  
157 above except the DCM/crude oil mixture was not added. Three replicates of each sample were  
158 prepared to capture variability. The composition of the different nanoparticle formulations investigated  
159 are listed in Table 1. All batch experiments were performed in triplicate.

## 160 Analyses

161 Aqueous nanoparticle samples were acid digested in 6 mol/L HCl and then diluted by a factor of 10 with  
162 1 mol/L HCl prior to analysis by ICP-OES (Prodigy) to quantify total iron (method detection limit (MDL) of  
163 4  $\mu\text{g/L}$ ). Nanoparticle mass bound on the OSS or the SS ( $\mu\text{g}$  of NP/dry g of sand) was estimated from the  
164 difference between the initial and final (after 48 h of exposure) nanoparticle concentration.

165 High Resolution Transmission Electron Microscopy (HR-TEM) analysis was performed using a JEOL 2010F  
166 with an acceleration voltage of 200 keV. The nanoparticles were dispersed in ethanol, sonicated, and a  
167 droplet placed on a holey carbon coated Cu grid, which was allowed to air dry before being analysed.

168 Dynamic Light Scattering (DLS) was used to characterize nanoparticle hydrodynamic size and uniformity.

169 A subsample (3 mL) of the nanoparticle suspension in hexane (prior to Pluronic coating) or in water  
170 (after Pluronic coating) was placed into a polypropylene cuvette and analyzed by DLS (Brookhaven  
171 90Plus Particle Size Analyzer).

172 Sample magnetization was measured using a Vibrating Scanning Magnetometer (VSM; Quantum Design  
173 MPMS 3). Sand samples (as described above) were packed into polycarbonate VSM sample holders.  
174 VSM was performed at a temperature of 70 K cooled by liquid N<sub>2</sub> to prevent material movement within  
175 the samples and was performed with a maximum magnetic field strength of 1 T. Magnetic properties of  
176 nanoparticles required for quantitative analysis were determined using freeze-dried Formulation A  
177 particles prepared using a Labconco Freezone 2.5 freeze drier.  
178 X-Ray CT samples (as described above) were packed into 4.5-mL polypropylene cuvettes and scanned  
179 with an open directional high-power micro-focus X-ray tube (240 kV, GE Phoenix v|tome|x m compact  
180 micro CT system) equipped with a GE DXR detector array. X-ray scan parameters (Table S1) were chosen  
181 to ensure optimal X-ray penetration and contrast. An acceleration voltage of 80 kV with a beam current  
182 of 80  $\mu$ A resulted in strong visual contrast between iron oxide and silica materials. Raw images were  
183 processed using Volume Graphics software (VGStudio Max) as two separate materials (iron oxide and  
184 silica) using beam hardening factors between 9.5 and 9.6. Both OSS and SS sample images were  
185 processed identically.

## 186 **Results and Discussion**

### 187 **Nanoparticle Design**

188 Engineered nanoparticles were designed to facilitate stability in aqueous suspension and favourable  
189 interactions with a hydrophobic phase such that particles would bind to the oil-water interface while  
190 avoiding adsorption directly to mineral soil material. To achieve this behaviour, an amphiphilic polymer  
191 coating was applied to a hydrophobic nanoparticle core (Scheme 1). The nanoparticle core was  
192 composed of iron oxide produced via a co-precipitation of iron salts, and made hydrophobic by oleic  
193 acid chemisorbed to the iron oxide surface through monodentate chelation of its carboxylate group with  
194 iron surface atoms (Wilson and Langell, 2014). This oleic acid layer allows further functionalization by the

195 amphiphilic polymer which is bound to the nanoparticle by hydrophobic forces. The final particle  
196 structure allowed favourable thermodynamic interactions between hydrophobic segments of the  
197 amphiphilic polymer coating and a NAPL phase, causing nanoparticles to partition to the water/NAPL  
198 interface(Saleh et al., 2006, 2005; Wang and Acosta, 2013). Pluronic, an inexpensive, commercially  
199 available polymer with highly tunable hydrophobic/hydrophilic properties, was selected as the coating  
200 material. Pluronic is a block co-polymer composed of a middle, hydrophobic segment of poly-propylene  
201 oxide (PPO) and two outer, hydrophilic segments of poly-ethylene oxide (PEO). The Pluronic coating was  
202 attached to the iron oxide core through hydrophobic interactions between the oleic acid nanoparticle  
203 capping agent and PPO segments. The PEO segments of the coating extend into the aqueous phase  
204 surrounding the nanoparticle and prevent nanoparticle aggregation through steric stabilization. Details  
205 regarding the properties of different Pluronic polymers used are given in Table 2. It should be noted that  
206 a similar nanoparticle structure using Pluronic F127 has been previously reported in the literature for  
207 application in promoting cellular uptake(Lin et al., 2009), however findings in this paper indicate that  
208 F127 may not be the most appropriate polymer to promote targeted binding to viscous hydrocarbons,  
209 necessitating careful selection of amphiphilic surface coating molecules based on the desired  
210 application. L62 and L121 were investigated as more hydrophobic polymers having approximate relative  
211 hydrophobic/hydrophilic mass ratios ( $M_{n(\text{PPO})}/M_{n(\text{PEO})}$ ) of 4 and 9, respectively. P104 was investigated as a  
212 more hydrophilic polymer having an approximate relative hydrophobic/hydrophilic mass ratio of 1.5.

### 213 **Nanoparticle Characterization**

214 HR-TEM data showed that the nanoparticle iron oxide cores were spherical with an average diameter of  
215  $6.4 \pm 1.8$  nm (Figure 1 A), consistent with DLS measurement of particle cores in hexane – 8.4 nm  
216 diameter with a polydispersity index (PDI) of 0.260 (Figure S1). After Pluronic coating, the nanoparticle  
217 average diameter increased significantly to  $120.9 \pm 47$  nm (Figure 1 B), again supported by a DLS  
218 hydrodynamic diameter of 89.8 nm with a PDI of 0.138. This increase in size is attributed to the

219 formation of a miniemulsion of multiple iron oxide cores within a surfactant polymer coating(Cui et al.,  
220 2007; Landfester, 2000; Xu et al., 2006). These aggregates form due to the requirement for sufficient  
221 surface coverage with surfactant to realize a critically-stable state; smaller particles require higher  
222 surfactant concentrations(Landfester, 2000). Although aggregation increased with polymer coating, the  
223 dispersity of the particles remained narrow (Figure S1), indicating that aggregation was limited and large  
224 aggregates ( $> 1 \mu\text{m}$ ) were not present. HR-TEM images (Figure 1 C, D) demonstrated the crystallinity of  
225 the iron oxide particles individually and within an aggregate structure as indicated by the ordered rows  
226 of atoms. In contrast to the order seen within the iron oxide particles, the disordered fringe adjacent to  
227 the crystalline region of the nanoparticle observed in Figure 1D is interpreted as an amorphous polymer  
228 coating. The thickness of this coating was measured to be  $1.1 \pm 0.17 \text{ nm}$  which compares similarly to the  
229 thickness of Pluronic species adsorbed on hydrophobic surfaces through hydrophobic interactions  
230 (Brandani and Stroeve, 2003).

### 231 **Nanoparticle Binding**

232 The iron concentration in the filtered solution for nanoparticle Formulation A after exposure to OSS was  
233 significantly lower than the same formulation exposed to SS (Figure 2). Compared to the iron  
234 concentration of the initial solution ( $26.2 \pm 0.8 \text{ mg/L}$ ), a reduction of  $2.9 \pm 1.7 \text{ mg/L}$  (11.2%) was  
235 observed in the SS solution, and a reduction of  $14.8 \pm 0.84 \text{ mg/L}$  (56.4%) was observed in the OSS  
236 solution, indicating preferential nanoparticle adsorption to sand containing NAPL PHC. The estimated  
237 mass of nanoparticles bound to the sand after exposure was  $1.95 \pm 1.15 \mu\text{g/g}$  on SS, and  $7.89 \pm 0.56$   
238  $\mu\text{g/g}$  on OSS. Nanoparticle binding using Formulation A nanoparticles also exceeded binding observed  
239 under identical conditions using oleic acid-stabilized particles in the absence of Pluronic (Figure S2)  
240 indicating that Pluronic coating was important to the adsorption mechanism. When the particles are  
241 stabilized using only oleic acid, a double layer of oleic acid forms around the particles, creating a  
242 hydrophilic anionic carboxylate shell preventing interaction with the NAPL PHC. Wang et al. have

243 previously shown that addition of a cationic surfactant to an oleate nanoparticle coating system can  
244 significantly improve targeted binding to trichloroethylene by disrupting the charge of the bilayer(Wang  
245 and Acosta, 2013). A similar likely occurred in this system whereby Pluronic disrupts the oleate bilayer,  
246 allowing hydrophobic interaction between the oleate surface and the hydrocarbon/water interface.  
247 Furthermore, literature demonstrates that nanoparticle core properties influence particle adsorption  
248 behaviour by affecting coating conformation(Hotze et al., 2014). By layering Pluronic on top of oleic acid,  
249 we may lessen the effect of the core material on Pluronic conformation, leading to higher applicability of  
250 this coating strategy to various nanoparticle core materials. At this point, the mechanism of nanoparticle  
251 binding is not fully understood, but is hypothesized to be due to hydrophobic forces from the Pluronic  
252 coating, oleate coating, or a combination of both. Any nanoparticle binding to clean sand was attributed  
253 to van der Waals forces between the PEO segment of the Pluronic coating and the surface of the SiO<sub>2</sub>  
254 sand grains. Previous literature suggests hydrogen bonding occurs between the ether oxygen of PEO  
255 and the hydroxyl species on the surface of an acidic metal oxide such as SiO<sub>2</sub>(Addai-Mensah, 2007). This  
256 interaction necessitates that the PEO segments of the Pluronic coating should be short to maintain a  
257 strong binding contrast between clean and oil-impacted sands.

258 Direct, quantitative magnetic analysis of dried OSS before and after exposure to Formulation A  
259 nanoparticle solution was assessed using VSM. The saturation magnetization of solid Formulation A  
260 nanoparticles (required to calculate nanoparticle mass deposition in the sand sample) was measured  
261 using freeze-dried nanoparticles (Figure S3) and found to be 59.76 emu/g. The magnetic hysteresis  
262 curves for the magnetization of OSS samples as an applied magnetic field is cycled between polarities  
263 are shown in Figure 3. The saturation magnetization represents the maximum, steady-state  
264 magnetization reached by a sample under a strong magnetic field. The difference between the  
265 saturation magnetization in the OSS Control sample and the OSS+NP sample was attributed to the  
266 presence of magnetic nanoparticles. This difference was divided by the saturation magnetization of

267 Formulation A nanoparticles to give a mass concentration estimate (g/g) for bound nanoparticles of 8.13  
268  $\pm 0.27 \mu\text{g/g}$ . This VSM estimate compares closely to the estimate of  $7.89 \pm 0.56 \mu\text{g/g}$  determined  
269 indirectly by the decrease in iron concentration in the aqueous phase.

270 As the Pluronic hydrophobic/hydrophilic ratio of the nanoparticle coating increased (Table 1), the  
271 nanoparticle concentration in aqueous suspension decreased when exposed to OSS, implying greater  
272 nanoparticle binding (Figure 4). When compared to Figure 2, the smaller %NP decrease observed for  
273 samples in Figure 4 is primarily due to higher initial concentrations used in this binding experiment  
274 (estimated NP binding, however, is increased). The results in Figure 4 indicate that nanoparticle binding  
275 towards viscous hydrocarbons, specifically heavy crude oil, could be tuned by changing the surface  
276 coating formulation. These results are consistent with the behaviour of Pluronic-coated magnetite  
277 nanoparticles employed in biomedical applications - previous work using such particles as an MRI  
278 contrast agent for tumor imaging reported that Pluronic coating formulation could be adjusted to avoid  
279 adsorption to proteins and that a balance between the hydrophobic and hydrophilic characteristics of  
280 the polymer is critical to maintaining stability and dispersion (Jain et al., 2009). The trend observed in  
281 Figure 4 relating coating hydrophobicity to nanoparticle binding supports the theory that binding is due  
282 to hydrophobic interactions as proposed by Saleh et al. (Saleh et al., 2005). While the modification of  
283 the particles in this work is different, the interactions which lead to NAPL targeting behaviour remain  
284 the same. The larger, longer hydrophobic portions of the surface coating remain contracted in water  
285 and swell in the presence of oil, resulting in a more energetically favourable configuration of the particle  
286 at the oil/water interface and facilitate targeted binding behaviour.

### 287 **X-Ray CT Visualization of Nanoparticle Binding**

288 SEM characterization of sand following nanoparticle exposure was initially investigated as a direct  
289 method to support nanoparticle binding, but it was found that SEM provided insufficient magnification  
290 and resolution to observe nanoparticles on the sand surface (Figure S4). SEM/EDS (Energy Dispersive X-

291 ray Spectroscopy) was also investigated for the same purpose (Figure S5), but large sampling areas,  
292 restricted magnification and resolution, and overwhelming elemental presence of carbon and silicon  
293 restricted use of this technique, prompting investigation into alternative characterization methods.

294 X-ray micro CT was used as a qualitative tool to visualize the presence of nanoparticles bound to dried  
295 OSS and SS (as described previously). A set of 2D projection images was acquired during a full rotation of  
296 the sample, which was then used to reconstruct a 3D image consisting of volumetric pixels (voxels) using  
297 a filtering back-projection algorithm (Feldkamp et al., 1984). Micro-focus tubes defocus the electron  
298 beam proportionally to the electric power to prevent the sample from thermal destruction (Gusenbauer  
299 et al., 2016). The voxel size estimated by the GE Phoenix X-ray CT machine was determined to be 27.3  
300  $\mu\text{m}^3$  for all scanned samples.

301 The detection of a single iron oxide nanoparticle or nanoparticle aggregate within a voxel resolution  $>1$   
302  $\mu\text{m}^3$  is challenging. This detection method relied on the overall X-ray absorbance of many iron oxide  
303 nanoparticles within one voxel to differentiate a response from the absorbance of silica sand, and hence  
304 there was a concentration threshold of nanoparticles for which a positive detection can be determined  
305 within a single voxel. Based on preliminary studies it was determined that the detection capabilities of  
306 the micro CT system with the X-ray scan parameters used could be as low as 1 mg/kg nanoparticles in  
307 silica sand (Figures S6 to S8).

308 Formulation A nanoparticle suspension was selected for CT investigation based on the observation that  
309 binding improved with coating hydrophobicity (see Figure 4). The bottom-half of the cuvette shown in  
310 Figure 5(A-C) was packed with OSS only, while the remaining top-half of the cuvette was packed with  
311 OSS that had been exposed to the Formulation A nanoparticle suspension for 48 hours. For baseline  
312 comparison, the bottom-half of the cuvette shown in Figure 5(D-F) was packed with SS only, and the  
313 top-half of the cuvette was packed with SS that had been exposed to the Formulation A nanoparticle

314 suspension for 48 hours. The packing of two materials in a single cuvette allowed for direct comparison  
315 of each sample to its relevant control.

316 Blue colour has been used in Figure 5 to represent regions of high X-ray absorption (higher nanoparticle  
317 density) while green has been used for regions of lower X-ray absorption (lower nanoparticle density)  
318 within each cuvette. Due to the concentrations of iron oxide nanoparticles used in the binding studies,  
319 the threshold difference (between blue and green) was very narrow. As a result, there are sparse blue  
320 regions in the lower section of each cuvette which may occur due to variations in packing density. The  
321 images in Figure 5 do not depict absolute values of X-ray absorbance, but rather voxels which fall above  
322 (blue) and below (green) a uniform selected threshold to illustrate qualitative evidence of nanoparticle  
323 binding. All the images in Figure 5 were obtained using identical X-ray CT operating and image  
324 processing parameters so that comparative visual contrasts between Figure 5 (A-C) and (D-F) are  
325 possible. The increased X-ray absorbance in the top-half of the cuvette shown in Figure 5 (A-C) suggests  
326 substantial nanoparticle binding to the OSS. In contrast, the X-ray absorbance observed in Figure 5 (D-F)  
327 is minor for the SS that had been exposed to the nanoparticle suspension compared to Figure 5 (A-C),  
328 thus corroborating the indirect binding data (Figure 2).

## 329 **Conclusions**

330 Demonstrating nanoparticle targeted binding to NAPL PHCs from an aqueous suspension in a porous  
331 medium matrix is one of the critical elements of a targeted nanoparticle delivery system with potentially  
332 wide-reaching environmental implications. This work demonstrated that nanoparticle targeted binding  
333 to crude oil could be controlled as a function of surface properties determined by an amphiphilic  
334 polymer coating. Extension of this concept to the broader environmental monitoring and remediation  
335 field implies that targeted binding functionality could be applied to various nanoparticle core materials,  
336 potentially enhancing existing nanoremediation techniques and enabling the use of targeted contrast  
337 agents for advanced site characterization. Immobilization of engineered nanoparticles at the interface



338 between the target contaminant mass and water addresses key drawbacks of current *in situ*  
339 nanoremediation strategies, namely improving efficiency and cost,(Saleh et al., 2006, 2005) as well as  
340 limiting uncontrolled nanoparticle release(Tosco et al., 2014).

341 The application of X-ray CT to detect nanoparticles in a porous medium has great potential for further  
342 development. In this study, there are two notable implications to be drawn from X-ray CT visualization  
343 of nanoparticle binding. First, X-ray CT is introduced as a useful, non-destructive technique for quickly  
344 analyzing 3-D sand characteristics with minimal sample preparation. Second, it implies a useful  
345 application for nanoparticle targeted binding in porous media, that is, as a contrast agent for 3-D  
346 imaging of sands impacted with viscous heavy hydrocarbons such as crude oil. Based on these findings,  
347 it is conceivable that mobile nanoparticles may be visualized by X-Ray CT in dynamic transport studies.  
348 Such experiments may generate a wealth of data to further understanding of nanoparticle transport  
349 behaviour in porous media and characterize targeted nanoparticle binding in realistic hydrogeologic  
350 conditions. One current drawback of the technique as presented here is its qualitative nature, but with  
351 careful development of a proper analysis methodology, it may be possible to extract quantitative data  
352 from this technique. In a similar vein, the use of SQUID/VSM magnetic characterization introduces quick,  
353 non-destructive, quantitative analysis techniques with minimal sample preparation which may help to  
354 overcome difficulties with characterizing nanoparticle presence in sand samples.

355 The work presented here limits its scope to batch investigation and novel X-ray characterization as a  
356 proof-of-concept for a targeted binding strategy, but these findings open the door to larger scale  
357 experiments under more realistic conditions. The nanoparticle coating proposed here has demonstrated  
358 limited binding to clean silica sand implying that transport through porous media should be possible,  
359 however literature has demonstrated that interactions with natural macromolecules can change the  
360 surface of engineered nanoparticles, affecting their behaviour and fate in soils(Louie et al., 2016). To this  
361 end, research investigating the transport properties of these nanoparticles in various porous media, as

362 well as expanded application of this novel X-ray CT nanoparticle characterization technique is ongoing in  
363 pursuit of improving and demonstrating the application of this targeted delivery platform for  
364 nanoremediation and geophysical monitoring.

## 365 **Associated Content**

### 366 **Supporting Information:**

367 Additional characterization such as DLS to measure particle size in solution, SEM/EDS of clean/oily sand  
368 after nanoparticle incubation, and X-ray micro CT of NP-impregnated sand samples to examine  
369 thresholds as well as additional information on X-ray CT parameters are available free of charge via the  
370 Internet.

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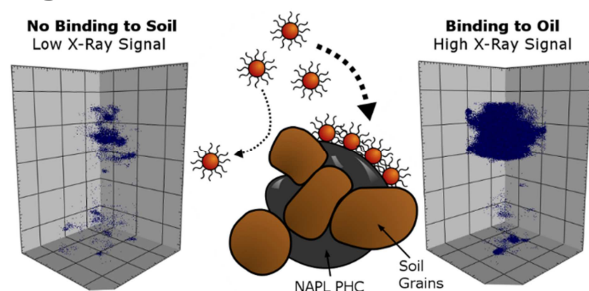
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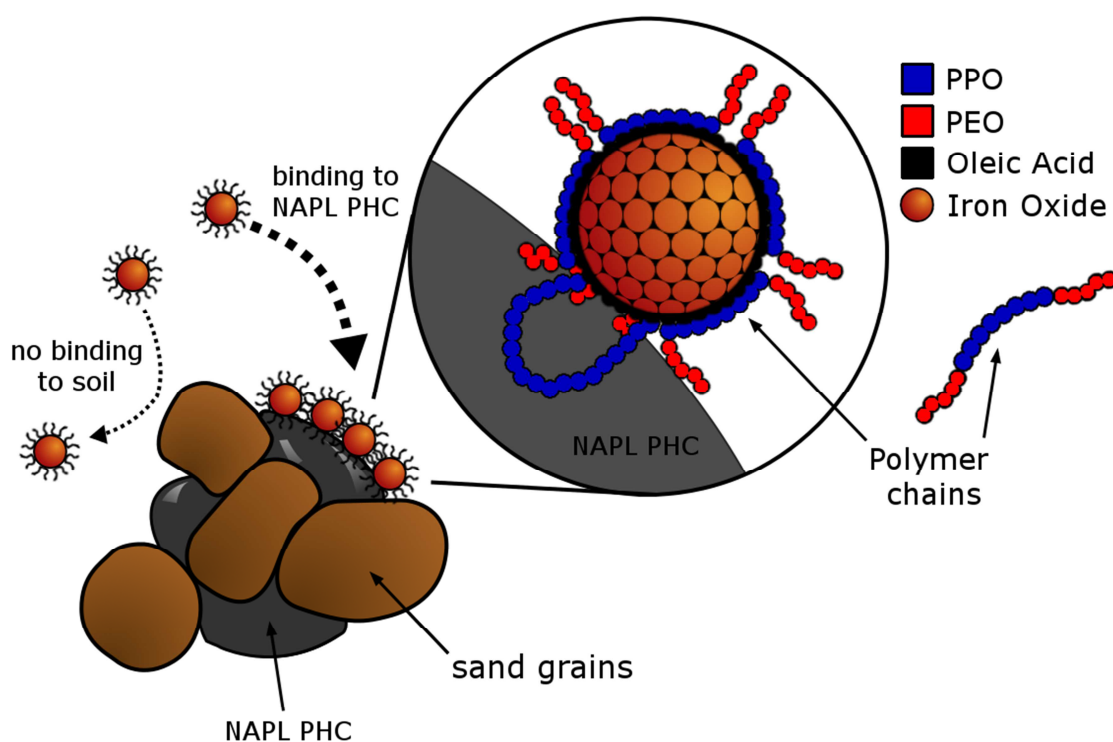
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581 **Figures & Tables**

582

583 **Graphical Abstract.** Engineered nanoparticles preferentially attach to NAPL PHC over soil grains. This is  
 584 observed through higher X-ray CT signal when NP are exposed to oil-impacted sand (right) compared to  
 585 clean sand (left).



586

587 **Scheme 1.** Schematic illustrating nanoparticles with an amphiphilic polymer coating (Pluronic) avoiding  
 588 adsorption to sand grains and specifically adsorbing to NAPL PHC through hydrophobic interactions.  
 589 Polyethylene Oxide (PEO; red) blocks of the coating extend into the aqueous phase and contract in the  
 590 NAPL PHC. Polypropylene Oxide (PPO; blue) blocks of the coating extend into the NAPL PHC and contract  
 591 in the aqueous phase. Oleic acid (black) binds the coating to the nanoparticle surface and interacts  
 592 favourably with the NAPL PHC when at the interface.

593 **Table 1.** Summary of nanoparticle formulations explored.

formulation ID <sup>a</sup>	Pluronic L62 (g/L) <sup>b</sup>	Pluronic L121 (g/L) <sup>b</sup>	Pluronic P104 (g/L) <sup>b</sup>	Dynamic Light Scattering diameter (nm)
A (A-100)	10	-	-	89.80
B-40	4	-	6	64.42
B-60	6	-	4	57.60
B-80	8	-	2	61.60
C-40	2	2	6	75.61
C-60	3	3	4	82.68
C-80	4	4	2	81.16

594 <sup>a</sup> Oleic acid is a common component of all formulations, present at a concentration of 0.1% v/v

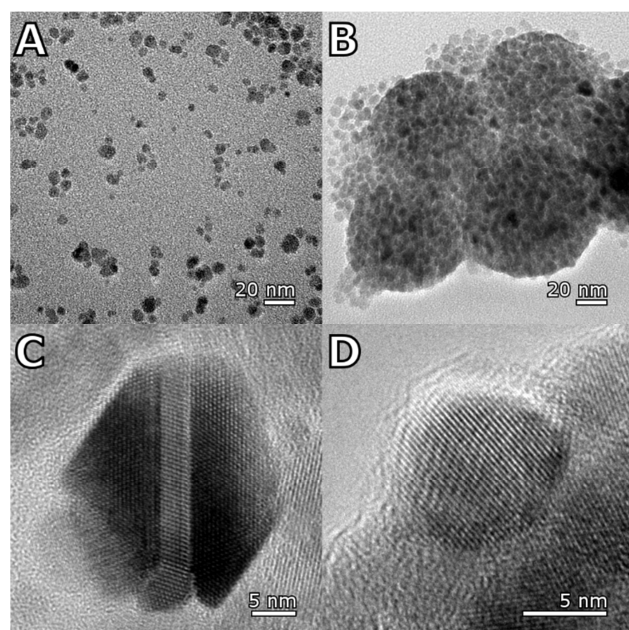
595 <sup>b</sup> concentration in aqueous solution prior to nanoparticle phase transfer.

596

597 **Table 2.** Properties of Pluronic polymers used for amphiphilic nanoparticle coating

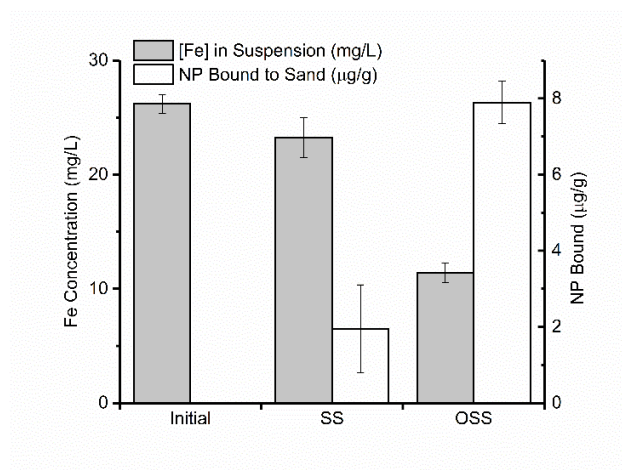
Polymer ID	M <sub>n</sub> (g/mol)	Structure	M <sub>n</sub> of PPO (g/mol)	M <sub>n</sub> of PEO (g/mol)	M <sub>n(PPO)</sub> /M <sub>n(PEO)</sub>
L62	2,250	PEO <sub>5</sub> -PPO <sub>31</sub> -PEO <sub>5</sub>	1,800	450	4
L121	4,000	PEO <sub>5</sub> -PPO <sub>62</sub> -PEO <sub>5</sub>	3,600	400	9
P104	5,000	PEO <sub>23</sub> -PPO <sub>52</sub> -PEO <sub>23</sub>	3,000	2,000	1.5

598



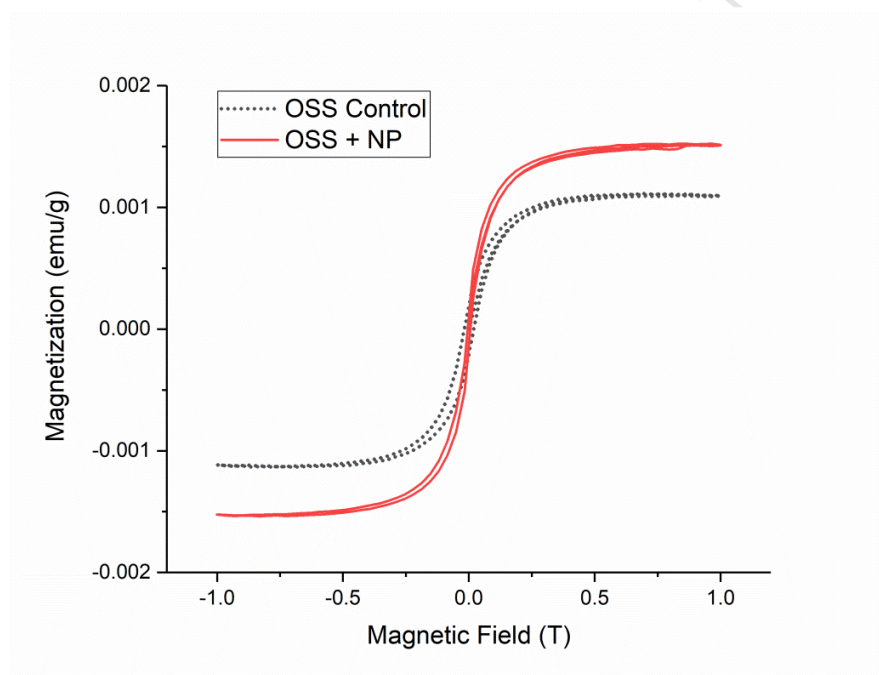
599

600 **Figure 1.** HR-TEM image of sample A-100: **A** Iron oxide nanoparticles prior to Pluronic coating, **B** Clusters  
 601 of iron oxide nanoparticles after Pluronic coating, **C** Iron oxide nanoparticle prior to Pluronic coating,  
 602 and **D** Edge of a Pluronic coated nanoparticle aggregate demonstrating crystalline nanoparticles within  
 603 an amorphous polymer coating.



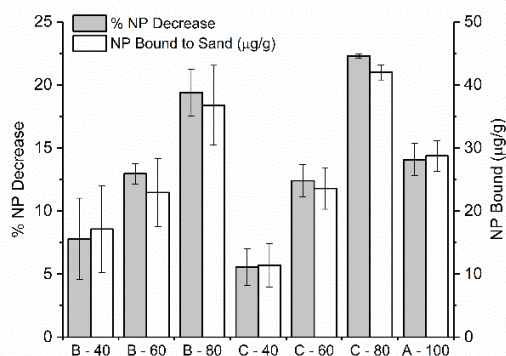
604

605 **Figure 2.** ICP-OES results demonstrating preferential binding of nanoparticle Formulation A to OSS when  
 606 compared to SS. Iron concentration in suspension was measured directly using ICP-OES. NP bound to  
 607 sand was calculated indirectly using the observed decrease of iron in the suspension. Error bars  
 608 represent  $\pm$  one standard error.



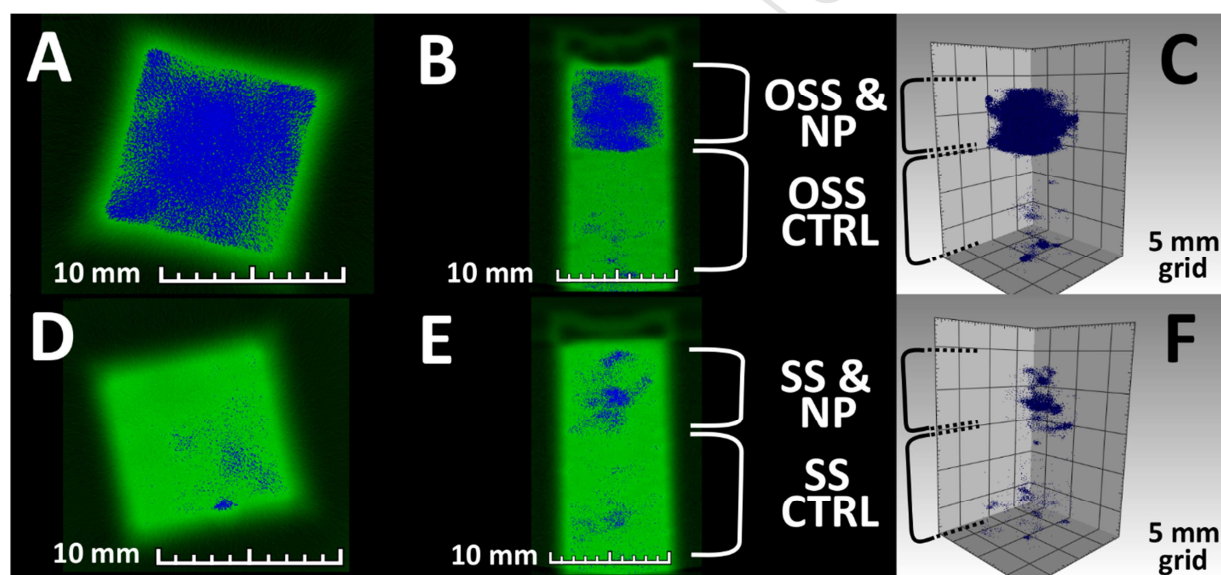
609

610 **Figure 3.** Magnetic hysteresis curves for OSS before (OSS Control) and after exposure to Formulation A  
 611 nanoparticle solution (OSS + NP). The saturation magnetization was  $1.037 \pm 0.016 \times 10^{-3}$  emu/g for the  
 612 OSS Control sample, and  $1.523 \pm 0.016 \times 10^{-3}$  emu/g for the OSS+NP sample.



613

614 **Figure 4.** Indirectly calculated nanoparticle (NP) binding increases as nanoparticle coating formulation  
 615 becomes more hydrophobic. The nomenclature X-# identifies nanoparticle samples (Table 1): X indicates  
 616 the types of Pluronic in the formulation (A: L62, B: P104+L62, C: P104+L62+L121), and # indicates the  
 617 percentage of the Pluronic in the formulation with higher relative hydrophobic content (L62 + L121).



618

619 **Figure 5.** X-ray Computed Tomography of Oil-impacted Silica Sand (OSS; **A**, **B**, and **C**) and Silica Sand (SS;  
 620 **D**, **E**, and **F**) samples. In each cuvette, the top portion contains sand contacted with Formulation A  
 621 nanoparticles for 48 h, filtered, washed, and dried while the bottom portion contains the respective  
 622 control (CTRL) sand, as indicated in **B** and **E**. **A** and **D** show the top view of each cuvette packed with OSS  
 623 and SS, respectively. **B** and **E** show the side view of each cuvette packed with OSS and SS, respectively. **C**  
 624 and **F** show a 3-D volumetric reconstruction of each cuvette for OSS and SS, respectively.

625

- Nanoparticles with polymer coating selectively adsorb to impacted sand
- Changing polymer coating composition influences adsorption behaviour
- Nanoparticles detected in impacted sand using novel application of X-Ray CT
- Targeted binding to crude oil in the presence of silica sand at ambient temperature

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